

A facile, rapid and sensitive detection of MRSA using a CRISPR-mediated DNA FISH method, antibody-like dCas9/sgRNA complex

Kyeonghye Guk^{a, b}, Joo Oak Keem^c, Seul Gee Hwang^{a, b}, Hyeran Kim^{a, b}, Taejoon Kang^{a, b}, Eun-Kyung Lim^{a, b*}, and Juyeon Jung^{a, b*}

^a Hazards Monitoring Bionano Research Center, Institute of Bioscience and Biotechnology (KRIBB), 125 Gwahak-ro, Yuseong-gu, Daejeon, 305-806, South Korea

^b Nanobiotechnology Major, School of Engineering, University of Science and Technology (UST), 217 Gajeong-rp, 34113, Daejeon, Republic of Korea

^c BioNano Health Guard Research Center, 125 Gwahak-ro, Yuseong-gu, Daejeon, 305-806, South Korea

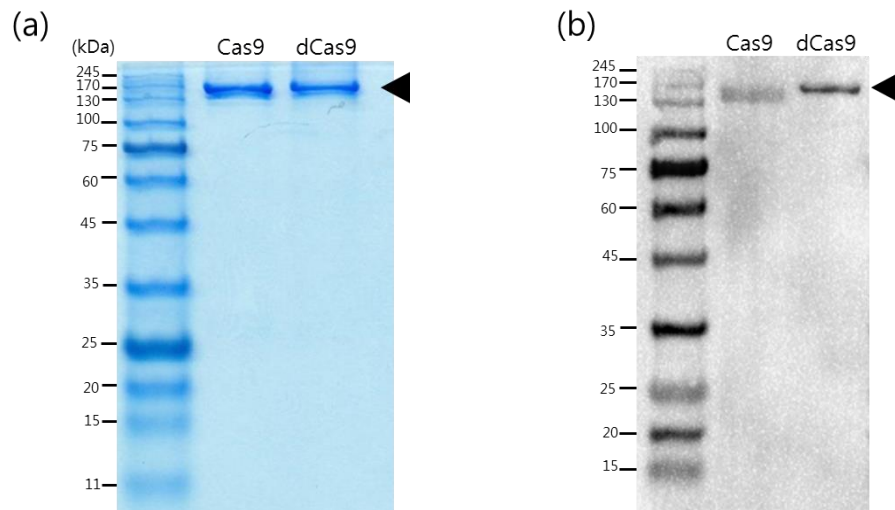


Figure S1. Identification of purified Cas9 and dCas9 by SDS-PAGE and Western blot analysis. (a) Purified proteins were applied to a 10% SDS-polyacrylamide gel under reducing condition followed by Coomassie Brilliant Blue staining. (b) Western blot using anti-6xHis monoclonal antibody and HRP-conjugated anti-mouse IgG Fc. The relative molecular weight (kDa) of commercial prestained markers is indicated on the left. The arrows indicate the purified Cas9 and dCas9.

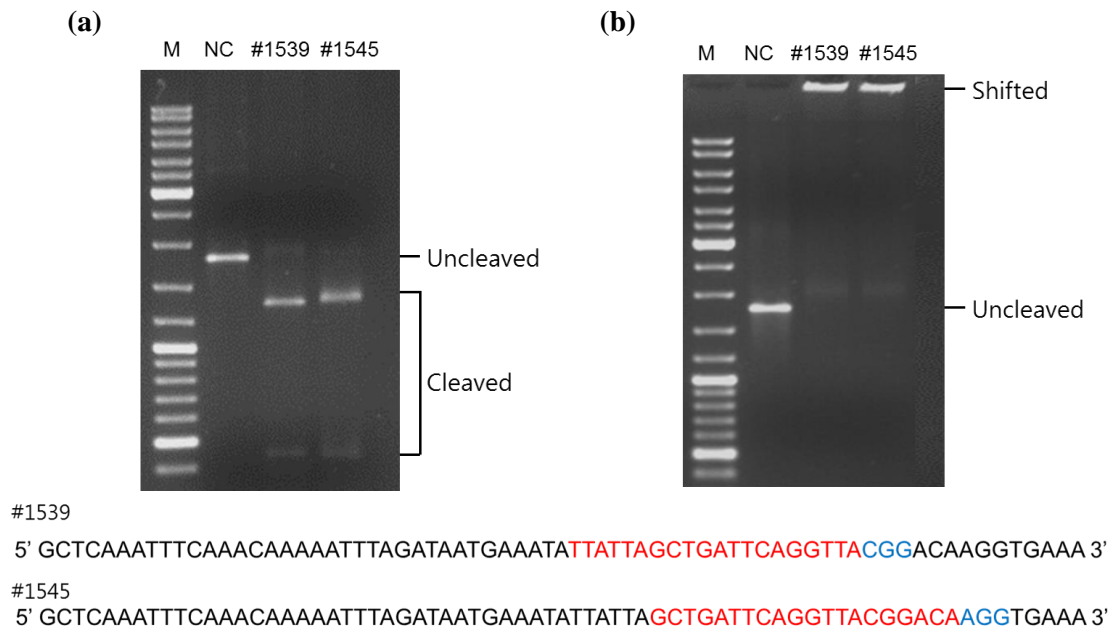


Figure S2. Site-specific DNA recognition of sgRNAs targeting the *mecA* gene in complexes with Cas9 or dCas9. The targeting efficiencies of two different sgRNAs (#1539 and #1545) were tested. (a) The Cas9/sgRNA-mediated cleavage of the *mecA* gene and (b) dCas9/sgRNA-mediated shift in mobility were monitored and visualized using 0.8% agarose gel electrophoresis. A negative control (NC) that lacked sgRNA was included for comparison. The sequences of the used sgRNAs are given under the graphs. The PAM and target sequences are indicated in blue and red, respectively.